



Implicit Motif Distribution based Hybrid Computational Kernel for Sequence Classification

Volkan Atalay^{1,2}, Rengul Cetin-Atalay^{1,3*}

¹Virginia Bioinformatics Institute, Virginia Polytechnic Institute and State University Blacksburg, VA 24061, USA, ²Department of Computer Engineering, Middle East Technical University, TR-06531 Ankara, Turkey, ³Department of Molecular Biology and Genetics, Faculty of Science, Bilkent University, 06533 Ankara, Turkey

ABSTRACT

Motivation: We designed a general computational kernel for classification problems that require specific motif extraction and search from sequences. Instead of searching for explicit motifs, our approach finds the distribution of implicit motifs and uses as a feature for classification. Implicit motif distribution approach may be used as *modus operandi* for bioinformatics problems that requires specific motif extraction and search, which is otherwise computationally prohibitive.

Results: A system named P2SL that infer protein subcellular targeting was developed through this computational kernel. Targeting-signal was modeled by the distribution of subsequence occurrences (implicit motifs) using self-organizing maps. The boundaries among the classes were then determined with a set of support vector machines. P2SL hybrid computational system achieved ~81% of prediction accuracy rate over ER targeted, cytosolic, mitochondrial and nuclear protein localization classes. P2SL additionally offers the distribution potential of proteins among localization classes, which is particularly important for proteins, shuttle between nucleus and cytosol.

Availability: <http://staff.vbi.vt.edu/volkan/p2sl>

Contact: rengul@bilkent.edu.tr

1 INTRODUCTION

Proteomics and systems biology are emerging fields as a result of recent advances in molecular biology, which produce large-scale genomic data. In addition, computational techniques generate new putative protein sequences predicted from genomes. Therefore, computational analysis techniques, which aim to develop tools for functional annotations to protein sequences, are highly required in the post-genomic era (Eisenberg *et al.*, 2000). Accurate and descriptive protein sequence annotations depend on the determination of particular amino-acid subsequence called protein motifs that are specifically related to a biochemical function. In general, computational motif dis-

covery tools (Bailey & Elkan, 1994, Blekas *et al.* 2003), focus on the explicit search and identification of motifs. Sequence motifs was identified by three approaches: deterministic patterns (PROSITE, PRINTS), profiles (BLOCKS, PROSITE, MEME) and probabilistic patterns with hidden Markov models (PFAM) (Henikoff and Henikoff 1991, Attwood and Beck 1994, Bailey and Elkan 1994, Bucher and Bairoch 1994, Bateman *et al.* 2004). Sequence motif discovery is more difficult than it might seem because many sequence motifs are discontinuous and not only the spacing between the residues but also the order in which they occur may vary. Therefore, whole or partial sequence analysis may be applied to each particular functional protein annotation problem. The sequence motifs are used for functional annotation of proteins with additional operations such as PROFILESCAN, BLIMPS and MAST, (Henikoff and Henikoff 1991, Bailey and Gribskov 1998,). The approach we present here, which is based on the use of distribution of implicit motif features may be employed as a general computational kernel for classification problems in genome and proteome analysis that necessitate specific motif extraction and search, which is otherwise computationally prohibitive.

Analysis of the subcellular localization of a newly identified protein is invaluable for the characterization of its function. Eventually, studying subcellular localization and developing predictors may have implications in systems biology in terms of a better representation of cellular machinery with respect to the site of protein action (Demir *et al.*, 2002). There have been several studies to predict protein subcellular localization based on primary sequence as well as three-dimensional structure information (Dierendonck *et al.*, 1997; Nakai, 2000; Emanuelsson, 2002, Mott *et al.*, 2002) Traditional computational predictors concentrate on the presence of signal peptide cleavage sites on protein sequences using machine-learning techniques (i.e. NNPSL, TargetP, SignalP, SortPred, SubLoc). Recently, in addition to machine learning techniques, hybrid methods that cover extensive biological knowledge

* To whom correspondence should be addressed.

such as protein sequence homology, protein and literature database text analysis, have been applied to attain a better accuracy of prediction (i.e. LOC3D, PA-SUB, PSORT-B, SMART) (Emanuelsson *et al.*, 2000, Mott *et al.* 2002 Nair& Rost, 2003; Gardy *et al.*, 2003; Lu *et al.*, 2004).

The aim of this study is to demonstrate the application of distribution of implicit motifs as a feature for classification on the prediction of subcellular localization of proteins in eukaryotic organisms. Therefore, we designed and developed a system called prediction of protein subcellular localization (P2SL). P2SL uses local subsequence features along with various amino acid similarity schemes. We used the self-organizing map for prototype feature extraction and implicit protein sorting signal (motif) distribution. Subsequently a set of support vector machines is used for the classification of the features extracted by SOM.

2 SYSTEM AND METHODS

2.1 Computational problem definition

From computational point of view, the prediction of subcellular localization among n -compartments can be stated as an n -class classification problem. In P2SL system n is determined by the following 4 compartments: ER targeted (all ER mediated membrane enclosed proteins), cytosolic, mitochondrial and nuclear:

given n sets of primary sequence samples that are drawn from n unknown probability distributions, construct a system able to learn to label a compartment (one of n) when used on new samples drawn from the same probability distributions.

Here, we assumed that the group of protein sequences targeted to a compartment has a particular probability distribution. However, neither the type nor the parameters of the distribution are known. From computational point of view we are given n sets of primary sequence samples each of which is drawn from one of the n above-mentioned groups. In addition we know from which group these samples are drawn; that is the samples are labeled with their correct compartments. Subsequently a new protein sequences (exclusive of the labeled sets) is classified to one of the n compartments. The problem is to construct a system that will predict to which of the groups the new sequence belongs. n -sets of labeled samples would be used for training and the trained system would be employed to predict the label of the new sample.

2.2 General approach

The system was composed of a set of modules (Fig.1). A module indicated whether the input sequence was of one

of two particular classes. Therefore, if there were n classes, we had $k=n(n-1)/2$ modules. Each module was composed of an encoder, a distribution extractor, and a decision maker (Fig.1A). The input to a module was amino acid primary sequences. An input primary sequence was decomposed into subsequences. Each subsequence was then encoded with an amino acid similarity matrix (see below). We fed the encoded subsequences to SOM. During the training phase of SOM, distributions of amino acid order for two classes particular to the module were determined by the winning nodes of SOM. In the space, the boundary between the two classes was learned by a SVM. In the test phase, when the subsequences of an unknown query sequence were given, according to the corresponding winning nodes in the SOM, a distribution was obtained and it was fed to SVM to be classified into one of the two already learned distributions

2.3 Self Organizing Maps and Support Vector Machines

We used two machine-learning methods in a hybrid way: self-organizing maps and support vector machines. SOM has a competitive unsupervised learning algorithm and it can be used for clustering and to construct nonlinear projection of the data onto a low-dimensional space as well (Kohonen, 1997). SOM algorithm leads to an efficient representation of the statistical structure implicit in the input samples and it generates a set of prototype vectors representing templates for a group of input samples mapped to a node or to a set of neighboring nodes. The latter process is called as vector quantization, which is a form of dimensionality reduction. We explore particularly this property of SOM. SVM is a discriminative supervised statistical machine learning method for regression and classification (Boser *et al.*, 1992; Burges, 1998; Vapnik, 1998; Scholkopf *et al.*, 1999; Cristianini and Shawe-Taylor, 2000). When learning from data, traditional machine learning and pattern recognition methods minimize training error in a particular norm. On the other hand, SVM is an approximate implementation of structural risk minimization and it constructs a hyperplane, i.e. linear division, in terms of a subset of the training data and the hyperplane separates negative and positive examples with the maximum margin. Prior to finding the hyperplane, it is possible to map, linearly or non-linearly, the training data into a higher dimensional space by a kernel function to seek for a linear boundary. In addition, SVM permits some misclassifications by building a soft margin, which can be controlled by a parameter. This is very helpful when data is noisy. After finding the hyperplane, previously unseen data in test set can be labeled by SVM. In practice, SVMs have been used in many real life applications and particularly, they have already been applied for

several problems in bioinformatics and computational biology (Noble, 2004).

2.4 Implicit motif frequency distribution by SOM and obtaining class prototype distribution

Extraction of implicit motif frequency distribution occurred during training of SOM and in this phase, subsequences of a subcellular localization class were topologically grouped and prototypes for each group were obtained. Therefore, the input data set could be represented by a set of SOM prototype vectors, which were denoted by the winning nodes in the SOM. The problem of finding motifs particular to a class may then be interpreted as finding the nodes, i.e. prototype motifs, specific to a class. However, it is very hard to determine these motifs and the nodes specific to a class when sequence motifs are discontinuous. In addition the spacing between the residues

and also the order in which they occur may vary. The projection of these signal patches to one-dimension disperses the motif information throughout the protein primary sequence. Therefore we use the distribution of winning nodes corresponding to each subsequence and the whole protein sequence is represented by the occurrences, i.e. distribution, of the winning nodes.

During the iterations, the prototype vector of each node is contributed by the input vectors mapped not only to the node itself but to the neighboring nodes as well. At the end of iterations, the prototype vectors are in fact local averages of subsequence data. Although the elements of input vectors are integers, the prototype vectors are composed of real numbers and therefore there may not exist one-to-one mapping back to amino acids subsequences from these prototype vectors that are the representatives of sequence motifs. Furthermore, we make use of the distribution of prototype vector occurrences rather than the

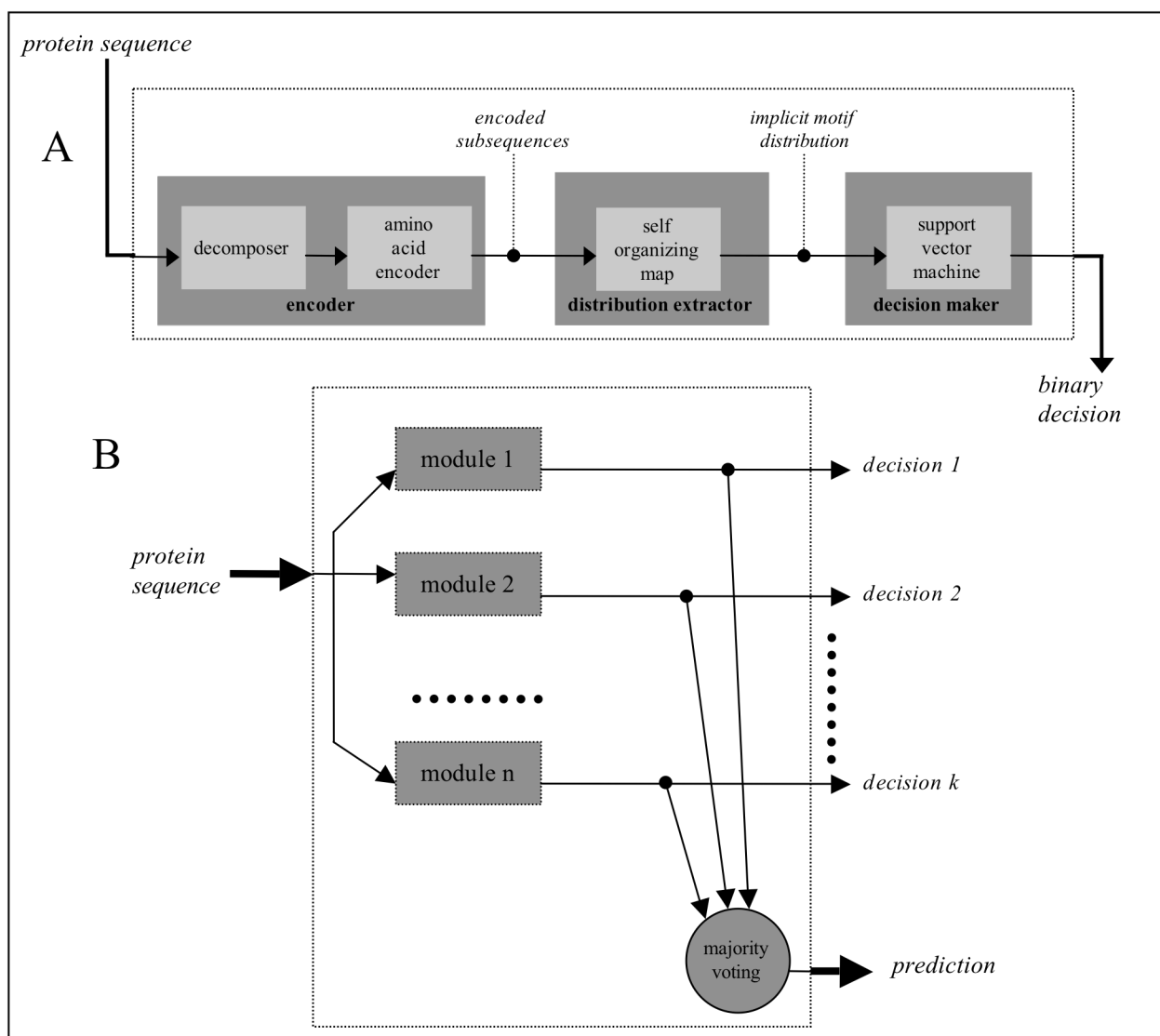


Fig. 1. Internal diagram of a module (A). General diagram of P2SL (B).

prototype motif vectors themselves.

2.5 Protein sequence encoding

Protein sequences are composed of letters corresponding to amino acids and these letters are non-numerical entities. In order to be able to perform computation on these sequences amino acids should be represented by numerical values. We used amino acid similarity matrices for this purpose and we called this process as encoding.

Amino acids were first encoded by the corresponding row found in amino acid similarity matrices. For example each amino acid in “MQT” subsequence could be encoded as follows $M=[-1\ 0\ -2\ -3\ -5\ -1\ -2\ -3\ -2\ 2\ 4\ 0\ 6\ 0\ -2\ 2\ -1\ -4\ -2\ 2]$, $Q=[0\ 1\ 1\ 2\ -5\ 4\ 2\ 1\ 3\ -2\ -2\ 1\ -1\ -5\ 0\ -1\ -1\ -5\ -4\ 2]$, $T=[1\ -1\ 0\ 0\ -2\ -1\ 0\ 0\ -1\ 0\ -2\ 0\ -1\ -2\ 0\ 1\ 3\ -5\ -3\ 0]$. Then, they were concatenated in the order that they are found in the sequence (MQT= $[-1\ 0\ -2\ -3\ -5\ -1\ -2\ -3\ -2\ 2\ 4\ 0\ 6\ 0\ -2\ 2\ -1\ -4\ -2\ 2\ 0\ 1\ 1\ 2\ -5\ 4\ 2\ 1\ 3\ -2\ -2\ 1\ -1\ -5\ 0\ -1\ -1\ -5\ -4\ 2\ 1\ -1\ 0\ 0\ -2\ -1\ 0\ 0\ -1\ 0\ -2\ 0\ -1\ -2\ 0\ 1\ 3\ -5\ -3\ 0]$) and a numerical subsequence vector was obtained. The effect of various encodings by amino acid similarity matrices are given and discussed in Section 3.3 and particularly in Table 1 by indicating the best and worst encoding amino acid similarity matrices.

2.6 Implicit motif frequency distribution

Let X denote an encoded protein sequence and that S denote a protein subsequence of length κ . If the dimension of SOM is $m*n$, any SOM node (i,j) can be represented as an integer via the map $f(i,j)=i*m + j$. Hence, a vector v of length $(m*n)$ can be associated with each protein sequence X . The element of v is initialized to 0. For each subsequence S , the corresponding winning node $w=(k,l)$, $v[f(w)]$ is incremented. When all the subsequences are processed, each entry of v is normalized by the total number of subsequences in X . v gives the distribution of winning SOM nodes for a protein which becomes the input data for classification. This step is given in the following algorithm.

```

for a protein sequence  $X$  do
  initialize each entry of  $v$  to 0
  for each encoded subsequence  $S$  do
    find the winning node  $w$  among the SOM
    nodes
    increment  $v[w]$  by 1
  end for
  for each component  $i$  of  $v$  do
    divide  $v[i]$  by total number of
    subsequences
  end for
end for

```

We used SOM_PAK: The Self-Organizing Map Program Package (Kohonen *et al.*, 1996) as a tool for SOM. The distributions were then used to train support vector ma-

chines. Test was consequently performed on the trained SVMs. We used SVMLight as a software tool (Joachims, 1999) based on one-against-one method.

2.7 Decision making: classification of class prototypes

In each module, we had a vector v at the output of SOM that represented the distribution of subsequences of an input sequence. These distributions were then used to train support vector machines to discriminate between the positive and negative class sequences. Test was consequently performed on the trained SVMs. SVMs were originally designed for binary classification. On the other hand, our problem was a multi-class classification problem. There are currently a few ways available to extend SVM to multi-class classification and we chose one-against-one method since it has been empirically proven to be more suitable for practical use (Hsu and Lin, 2002). In one-against-one method, if there are n classes, then $n(n-1)/2$ binary classifiers are built and each one is trained on data only from two classes. Classifier results were then combined by majority voting scheme in which the predicted class was the most voted one. Majority voting scheme for classifier combination has also been proven to be performing well for independent classifiers (Lam and Suen, 1994; Kittler *et al.*, 1998; Jain *et al.*, 2000).

2.8 Dataset

The dataset consisted of 3115 ER targeted (ER), 1780 cytoplasmic (C), 1148 mitochondrial (M) and 2225 nuclear (N) animal protein sequences derived from Swiss-Prot database. Major part of the sequence data was taken from PA-SUB (Lu *et al.*, 2004), in addition to manually selected sequences from Swiss-Prot database.

3 RESULTS

3.1 P2SL prediction outline

Our approach finds the frequency distribution of protein subsequences for each subcellular localization class and then uses this distribution as a feature for classification independent of explicit motifs. ER targeted, cytosolic, mitochondrial or nuclear class probability distributions were represented by samples of subsequence distributions over SOM. Distributions were extracted and learned during the training phase while prediction was performed in the test phase. The following binary SVM classifiers: ER versus C, ER versus M, ER versus N, M versus C, M versus N and N versus C were used. Each class was voted over three classifiers. Majority voting then gave the predicted class (Fig 1).

3.2 Selection of Subsequence Parameters

It has been reported that proteins which are to be ER targeted have a sorting signal called as signal peptide of length approximately 20-25 amino acids in the N terminal of the primary sequence while proteins destined to the mitochondria usually contain an N-terminal mitochondrial transfer peptide, on the average 35 amino acids long (Nakai, 2000; Emanuelsson, 2002). There is not an explicit signal sequence start site on entire protein sequence of cytosolic and nuclear proteins. Nuclear localization signals can be found at any part of the sequence whereas cytosolic proteins and some of the nuclear proteins don't carry any signal motif at all. Hence, for each subcellular localization prediction class, part of sequence to be examined (L), window length (κ), and slide offset for subsequences (t), were determined as different values after rigorous computational experimentation. In order to characterize the proteins targeted to ER, we analyzed the first 30 amino acids ($L=30$), with a window length of 5 ($\kappa=5$) and sliding the windows by one amino acid ($t=1$) at a time. Similarly, for the proteins targeted to mitochondria, we investigate the first 40 amino acids ($L=40$) with window length of 5 ($\kappa=5$) and sliding the windows by one amino acid ($t=1$). For proteins targeted to nucleus or destined to act in cytosol, our results demonstrated that analysis of the entire sequence ($L=all$), with a window size of 15 ($\kappa=15$) and sliding the window by 5 amino acids ($t=5$) gives good prediction accuracy in addition to its advantage in computational time complexity.

3.3 Amino acid encoding

Although, the most popular way of encoding of amino acids reported in the literature is to represent each amino acid in binary form by sparse encoding, this representation ignores the context information. There has been a large amount of research to characterize the physico-chemical and biochemical properties of individual amino acids and assign them numeric values (Kawashima *et al.*, 1999). In our study, amino acids were represented numerically by the similarity between them based on biochemical similarity propensities to each other by using amino acid similarity matrices (see Methods). These matrices are currently being used for scoring of protein sequence alignments. We included 73 different amino acid similarity matrices into our study and experimentally determined best amino acid sequence encoding matrices for each subcellular class. Table 1 presents list of best and worst amino acid encoding matrices used during subcellular class prediction.

Table 1. Best (A) and worst (B) encoding amino acid similarity matrices for ER, C, M and N protein sequences. (X) indicates the best and the worst encoding matrices for ER, C, M and N protein sequence classes.

A				
Best encoding amino acid similarity matrices	ER	C	M	N
BLOSUM50 substitution matrix	X	X	X	
BLOSUM80 substitution matrix				X
Log odds matrix for 40 PAMs		X	X	
Residue replace ability matrix	X	X	X	X
Structure-based comparison tables for alpha or beta helix class		X		
The 250 PAM transmembrane protein exchange matrix	X	X		
The PAM-120 matrix			X	X
B				
Worst encoding amino acid similarity matrices	ER	C	M	N
Conformational similarity weight matrix	X		X	X
Context-dependent substitution matrices for alpha helix or turn or coil	X	X	X	X
Context-dependent substitution matrices for buried beta or turn or coil	X	X	X	X
Secondary structure from multiply aligned sequences			X	X
Structure-derived correlation matrix	X	X	X	X
Substitution matrix from spatially neighboring residues			X	X
WAC matrix constructed from amino acid comparative profiles		X	X	X

3.4 SOM and SVM Parameters

There are not so many analysis studies on SOM (Vesanto and Alhoniemi 2000, Kostiainen and Lampinen 2002, Kohonen 1997, Luttrell 1994) and only a few guidelines exist on its practical use. One of the most important parameters in SOM algorithm is the choice of map size. Wang *et al.* recommends to use $5*\sqrt{N}$ where N is the number of training patterns input to SOM (Wang *et al.* 2002). After performing intensive experiments, the good performing map sizes came out to be in the range $[\sqrt{N}, 5*\sqrt{N}]$. The map sizes that were finally adapted are as follows: ER (versus C, M, N) classifiers 47x40, M (versus C, N) classifiers 39x33 and N (versus C) classifier 55x47. Rectangular grid was chosen for all of the SOMs. Training was performed in two phases. First, the prototype vectors of the map units were ordered. During the second phase, the values of the prototype vectors were fine-tuned. In ordering, the neighborhood radius was taken as $(0.8*\text{maximum of the map size})$ and decreased to 1 while the learning rate decreased to zero. In fine-tuning, the neighborhood radius started with $(0.2*\text{minimum of the map size})$ and again decreased to 1. Learning rates were 0.1 and 0.01 for ordering and fine-

tuning, respectively. The number of iterations in each phase was related to the number of training patterns. In ordering, it was 1 or 2 times of the number of training patterns while in fine tuning it was 3 or 4 times. We did not normalize the input data to SOM since the encoding inherently brought some information and this might have been removed by normalization.

We used the following combination of binary SVM classifiers: ER versus C, ER versus M, ER versus N, M versus C, M versus N and N versus C. The major reason in the choice of the above binary classifiers was that during our previous experiments with "one versus rest" type of SVM classifiers, the classifiers were ranked as ER versus (C,M,N), M versus (ER,C,N), N versus (ER,C,M) and C versus (ER,M,N) in terms of the classification accuracy. That is why we have chosen first all combinations of ER then other possible combinations of M and the last possible combination N versus C.

Radial basis function kernel performed better than the other kernels for all of the classifiers. In this case, the values of two parameters should be chosen: parameter c , which determines the soft margin and γ that is the inverse width of the basis function. We simply did a grid search in order to find the values of these parameters. For ER and M classifiers $c=2.0$ and $\gamma=0.6$ while for N classifier $c=1.0$ and $\gamma=0.1$. We have observed that although SVM performance was affected by the changes in these parameters the overall prediction rate remains stable. The number of support vectors of the individual classifiers after training ranged between 1380 and 2370.

3.5 4-fold cross validation and performance assessment of P2SL

In order to assess the performance of the P2SL system, we performed 4-fold cross validation. Overall accuracy of the system was 81.96% on the average with a standard deviation of 0.43. However, overall accuracy does not represent the true performance of prediction methods in general. Therefore further statistical measures are required for detailed analysis of prediction. The results given by the confusion matrix were obtained by the average of 4-fold cross validation experiments (Table 2).

In addition to confusion matrix for each class the evaluation of the P2SL prediction results was presented by four statistical measures defined as: precision, recall, F-score and specificity. These statistical measures were given in terms of true positives (tp, number of patterns predicted as of class K and actually they are of class K), false positives (fp, number of patterns predicted as of class K and they belong actually to other classes.), true negatives (tn, number of patterns predicted not to be of class K and they actually do not belong to class K) and false negatives (fn,

number of patterns predicted not to be of class K and they actually belong to class K).

Table 2. Confusion matrix indicating the average percentage values of 4-fold cross validation over the test set. Standard deviations were between 0.34 and 2.62. Test sets had 1780 ER targeted, 445 cytosolic, 287 mitochondrial and 890 nuclear proteins, which were mutually exclusive of the training sets each time.

Actual	Predicted label			
	ER	C	M	N
ER	85.63	6.19	3.29	4.89
C	2.36	79.33	3.65	14.66
M	5.66	7.23	83.80	3.31
N	1.43	19.94	3.29	75.34

The employed statistical measures are then given in terms of the above definitions.

$$\text{precision} = \frac{tp}{(tp+fp)}$$

$$\text{recall} = \frac{tp}{(tp+fn)}$$

$$F\text{-score} = 2 * \text{precision} * \text{recall} / (\text{precision} + \text{recall})$$

$$\text{specificity} = \frac{tn}{(tn+fp)}$$

Table 3 shows the average of these statistical measures for 4-fold cross validation. Precision measured the portion of the correctly assigned subcellular classes. Prediction precision of cytosolic proteins is relatively low, because there was not a particular localization signal in proteins function in cytosol, whereas subcellular localization of proteins having subcellular targeting peptides (Table 3 line 1 ER, M, and N) was predicted with high precision. On the other hand, recall statistics of P2SL that is the portion of the correct subcellular classes that were assigned, for all classes are similar, in addition to F-score, which was an even combination of precision and recall. Specificity, which is the ability of P2SL to correctly identify the proteins that do not belong to the class to which they are not targeted in vivo, was high for all classes.

Table 3. Performance assessment of the P2SL. The numbers indicate average percentage values of 4-fold cross validation. Standard deviations were between 0.29 and 2.62.

	ER	C	M	N
Precision	97.47	53.37	69.98	80.57
Recall	85.63	79.33	83.80	75.34
F-score	91.17	63.80	76.20	77.86
Specificity	97.56	89.57	96.66	93.56

3.6 Comparison with TargetP

TargetP is a neural network based tool for subcellular location prediction of proteins ((Emanuelsson *et al.*, 2000). Since TargetP is a computational tool similar to P2SL, we compared P2SL with TargetP. TargetP gives predictions in terms of only 3 classes: ER targeted (ER), mitochondria (M) and other. Therefore, we arranged the predicted labels of P2SL such that cytosolic (C) and nuclear (N) were in the “other” class. We compared the results of TargetP with P2SL only on the first set of 4 fold cross validation. The confusion matrices and statistical measures for two systems are given in Table 4A and 4B, respectively. On this set, the overall accuracy of P2SL was 88.83%, which was better than that of TargetP (82.54%). Similarly, all of the statistical measures of P2SL were better than those of TargetP. In addition, P2SL predicts for proteins targeted to nucleus and gives distribution possibilities about proteins shuffle between nucleus and cytosol.

Table 4. Comparison of Confusion matrices (A) and statistical measures (B) of results for Target-P and P2SL over one of the sets of 4-fold cross validation.

A

Actual	Predicted label			
		ER	M	Other
ER	TargetP	82.02	0.73	17.25
	P2SL	84.66	2.98	12.36
M	TargetP	18.12	54.70	27.18
	P2SL	8.71	81.53	9.76
Other	TargetP	2.25	8.61	89.21
	P2SL	2.25	1.80	95.96

B

		ER	M	Other
precision	TargetP	94.68	55.09	75.57
	P2SL	96.48	75.24	83.78
recall	TargetP	82.02	54.70	89.15
	P2SL	84.66	81.53	95.96
F-score	TargetP	87.90	54.90	81.80
	P2SL	90.19	78.26	89.46
specificity	TargetP	94.95	95.89	81.37
	P2SL	96.61	97.53	88.00

We compared TargetP and P2SL on the same set of proteins that is exclusive of the training set of P2SL and the results were given in Table 4A as confusion matrices and in Table 4B as statistical measures. TargetP’s prediction accuracy degrades while P2SL maintains similar performances for all locations. We did further comparative analysis on both predictors. We particularly looked at the cases where one or both of the systems fail (Table 5). Based on our analysis we observed that the percentage of cases in which TargetP predicted erroneously when P2SL predicted correct was higher than the percentage of cases in which P2SL predicted erroneously and TargetP predicted correct (compare *fn* values: 6.18 versus 3.54, 31.71 versus 4.88 in and 8.54 versus 1.80 in Table 5)

Furthermore, when we go through the individual cases on the results, we observed that when both systems predict wrong, in most of the cases, they predict the same compartment.

Table 5. Comparison of prediction errors made by TargetP and P2SL for ER, M and other (C and N) targeted proteins. Values are given as percentages. *tp* indicates true positives while *fn* indicates false negatives

TargetP	P2SL						
		ER		M		Other	
		<i>tp</i>	<i>fn</i>	<i>tp</i>	<i>fn</i>	<i>tp</i>	<i>fn</i>
<i>tp</i>		78.48	3.54	49.83	4.88	87.42	1.80
<i>fn</i>		6.18	11.80	31.71	13.59	8.54	2.54

It is hard to compare the computational complexities of TargetP and P2SL since they are composed of several complex modules. However, it is clear that the most time consuming part is the training. Once the systems are trained, obtaining a prediction for a query sequence is rather fast for both of the systems.

3.7 Protein distribution among localization classes

Each SVM classifier that we used for ER versus C, ER versus M, ER versus N, M versus C, M versus N and N versus C, gave a decision. These decisions were an input for majority voting (Fig. 1). Each class was voted by three classifiers generating a possibility of one over three. The sum of votes indicated the potential of the protein located to that class. However, it is meaningful to analyze and further interpret the decision results. To look at how many classifiers vote for a localization class gave protein distribution possibility among localization sites. Such distribution tendencies were particularly interesting in the case of proteins that shuttle between nucleus and cytosol.

We selected eight proteins reported to move to nucleus and interfere with gene expression control upon cell signal (Table 6). Four of those proteins (β -catenin, PKC α ,

hexokinase1, BCL2) have major functions in cytosol whereas the other four (BRCA1, p53, NF- κ -B, E2F1) is primarily known as transcription factors (Schmalz *et al.*, 1998; Hoetelmans *et al.*, 2000; Fabbro *et al.*, 2003; Ahuatzil *et al.*, 2004). P2SL analysis remarkably could predict that first four proteins locate with a possibility of 3/3 in cytosol and a possibility of 2/3 to nucleus.

Table 6. P2SL predicted Distribution of selected proteins that shuttle between nucleus and cytosol.

Protein Name	Cytosol*	Nucleus*
β -catenin	3/3	2/3
PKC α	3/3	2/3
Hexokinase1	3/3	2/3
BCL2	3/3	2/3
BRCA1	2/3	3/3
p53	2/3	3/3
NF- κ -B p100/p49	2/3	3/3
E2F1	1/3	3/3

Possibility of localization in Cytosol or Nucleus over three-voted classifiers.

4 DISCUSSION

Recent scientific interest on molecular biology moves toward systems biology that requires detailed functional analysis of genome and proteome. However large-scale data obtained through these analyses should be confirmed by individual *in vivo* experiments. It is critical to select experiments to be done since each of them is costly in terms of time and economical resources. Therefore predictive preliminary tools and models for this selection is invaluable. Many proteins may have more than one function depending on where and when they operate during its cellular activities (Cyert, 2001, Henderson and Fagotto 2002, Burchett 2003). Prediction of the cellular behavior of a protein is one of the research interests *in vivo* and *in silico* that includes subcellular localization of the proteins. Biological processes are too complex to be modeled by a single approach. There are several approaches for the prediction of subcellular localization based on: sorting signals (TargetP, PSORT), amino acid composition (SubLoc), sequence homology (PA-SUB), genomic context with phyla analysis and domain projection (SMART) (Mott *et al.* 2002). Each method gives a prediction with certain reliability thus single or combination of methods can be applied to a particular biological problem for better predictions. In our study P2SL aimed to find out the protein-sorting subsequences for major subcellular localization classes: endoplasmic reticulum (ER) targeted, cytosolic, mitochondrial and nuclear. However, it was difficult

to search directly for those subsequences particularly when they are discontinuous. In addition the spacing between the residues and also the order in which they occur may vary in the case of nuclear proteins. Therefore we use the distribution of implicit motifs for each localization class based on both amino acid order and composition.

It was relatively simple to predict the localization of the proteins to ER or Mitochondria where the N-terminal signal peptides are determinants whereas for proteins targeted to nucleus or retained in cytosol, the prediction was difficult. In addition these proteins may function in both compartments. They may carry both type of sequence signals composed of small signal subsequence patches throughout the entire sequences. As mentioned in Section 3.2, the signal patches for proteins targeted to nucleus or destined to act in cytosol were modeled as subsequences of length of 15 amino acids. This means that there are 15^{20} different possible subsequence compositions exist to look for. This is the cost of direct search, which is intractable. In P2SL system we applied a heuristic: the use of implicit motifs where the motifs to be sought are not determined explicitly but extracted in a different format from the training data. This extraction performed by machine learning and pattern recognition methods. Theory and practice of these techniques have been improved significantly and they have been successfully applied to areas such as character, speech, face recognition (Özer *et al.*, 2001; Mohamed *et al.*, 2004) as well as biological data analysis (Baldi and Brunak, 2001; Mjolsness and De Coste 2001).

When applying machine-learning techniques to a particular problem, optimization and parameter selection based on the nature of the input data, is highly important for the prediction performance. Therefore with different *in silico* experiments we tested various parameters in order to attain the best prediction performance. The parameter values that were selected were stable. A small change in their values did not affect significantly the prediction results.

We also compared several amino acid similarity matrices for amino acid encoding. When matrices, generated based on amino acid replacements observed in closely related sequences, were used, the prediction accuracy of subcellular classes was better than those of the matrices generated based on structural properties of proteins (Table 1). Our observation is in correlation with subcellular localization prediction methods that use phyla and domain analysis based on genomic context (Mott *et al.*, 2002).

When tested on nuclear proteins in general subcellular localization predictors do not achieve high prediction rates. Many of them are transcription related proteins and they are kept in cytosolic pool. When necessary they are translocated to nucleus (Cyert, 2001, Burchett 2003). Moreover some of the nuclear proteins are partitioned between two compartments acting in both locations

(Henderson and Fagotto 2002). In general, protein sequences may reveal themselves mostly as targeted to stay in cytosol. Therefore in P2SL, proteins targeted to nucleus (19.94%) or stayed in cytosol (14.66%) were the most confused ones bilaterally. This is very consistent with the recent results by Scott et al. where 16% of human proteins are predicted to be multi-compartmental. These results were in correlation with the cellular behavior of many nuclear proteins transcription factors for instance p53 or β -catenin, which are kept in a cytosolic pool until a cellular signal activates and sends them into the nucleus (Cyert, 2001, Henderson and Fagotto 2002, Burchett 2003). Accordingly the high rate of confusion obtained by P2SL analysis of nuclear and cytosolic proteins reflected biological system behavior. Therefore we took advantage of the binary P2SL-SVM classifiers to attribute possibilities for being in two different compartments as tested and demonstrated with the proteins given in Table 5. The proteins given in this table are initially described to act in cytosol, later their novel function in nucleus is discovered. However, proteins whose function mainly related to transcription regulation were predicted to be found in nucleus with 3/3 possibilities by P2SL-SVM classifier. If a single decision is expected, classifier results could be combined by majority voting scheme in which the predicted class was the most voted one. Nevertheless, we believe that assigning only one subcellular localization to proteins may neglect other functions for those proteins that have multiple action sites. Therefore P2SL provides to its users possibilities of subcellular localizations in addition to prediction. P2SL system will be updated through periodical training with the addition of new experimental data. This will provide P2SL users to keep up with the new emerging information with respect to subcellular localization.

In this study, we demonstrated that implicit motif distribution approach gave a simple but yet successful solution to the problem of subcellular localization using a large protein dataset. Our approach may be used as a computational kernel for various other biological problems that requires specific motif extraction and search. In addition, modular decision using SVM classifiers inherently provide decision possibilities for each classification incident. Computational methods defining classification possibilities may provide new insights to unexplained activities of proteins or may support to attribute new activities to proteins along with in vivo experiments.

ACKNOWLEDGEMENTS

We would like to thank M. Ozturk (Bilkent University, Turkey) for his critical discussions on manuscript and M. Ozarar for initial technical assistance. This work was supported by the Turkish Academy of Sciences to R.C.A.

(RCA/TUBA-GEBIP/2001-2-3), and R.C.A. partial funding was also provided by TUBITAK (Turkey).

REFERENCES

- Ahuatzi,D., Herrero,P., de la Cera,T. and Moreno,F. (2004) The glucose-regulated nuclear localization of hexokinase 2 in *Saccharomyces cerevisiae* is Mig1-dependent. *J Biol Chem.* **279**, 14440-14446.
- Attwood,T.K. and Beck,M.E. (1994) PRINTS--a protein motif fingerprint database. *Protein Eng.* **7**, 841-848.
- Bailey T.L. and Elkan C., (1994) Fitting a mixture model by expectation maximization to discover motifs in biopolymers. *Proceedings of the Second International Conference on Intelligent Systems for Molecular Biology*, 28-36.
- Bailey,T.L. and Gribskov,M. (1998) Combining evidence using p-values: application to sequence homology searches. *Bioinformatics* **14**, 48-54.
- Baldi,P. and Brunak,S. (2001) *Bioinformatics: The Machine Learning Approach*. Second Edition, MIT Press.
- Bateman,A., Coin,L., Durbin,R., Finn,R.D., Hollich,V., Griffiths-Jones,S. Khanna,A., Marshall,M., Moxon,S., Sonnhammer,E.L.L, Studholme,D.J., Yeats,C. and Eddy,S.R. (2004) The Pfam Protein Families Database. *Nucleic Acids Research Database Issue* **32**, 138-141.
- Blekas,K. Fotiadis,D.I., and Likas,A. (2003) Greedy mixture learning for multiple motif discovery in biological sequences. *Bioinformatics* **22**, 607-617.
- Boser,B., Guyon,I. and Vapnik,V. (1992) An training algorithm for optimal margin classifiers. *Proceedings Fifth Annual Workshop on Computational Learning Theory I*, 144-152.
- Bucher,P. and Bairoch,A. (1994) A generalized profile syntax for biomolecular sequences motifs and its function in automatic sequence interpretation. *Proceedings 2nd International Conference on Intelligent Systems for Molecular Biology*, 53-61.
- Burchett,S.A. (2003) In through the out door: nuclear localization of the regulators of G protein signaling. *J Neurochem.* **87**, 551-559.
- Burges,C.J.C. (1998) A tutorial on support vector machines for pattern recognition. *Data Mining and Knowledge Discovery* **2**, 121-167.
- Cristianini,N. and Shawe-Taylor,J. (2000) *An Introduction to Support Vector Machines*, Cambridge University Press.
- Cyert,M.S. (2001) Regulation of nuclear localization during signaling. *J Biol Chem.* **15**, 20805-20808.
- Demir,E., Babur,O., Dogrusoz,U., Gursoy,A., Nisanci,G., Cetin-Atalay,R. and Ozturk,M. (2002) PATIKA: an integrated visual environment for collaborative construction and analysis of cellular pathways. *Bioinformatics* **18**, 996-1003.
- Dierendonck,J.H., Bahar,I., Atilgan,A.R., Jernigan,R.L. and Erman,B. (1997) Understanding the recognition of protein structural classes by amino acid composition. *Proteins* **29**, 172-185.
- Eisenberg,D., Marcotte,E.M., Xenarios,I. and Yeates,T.O. (2000) Protein function in the post-genomic era. *Nature* **405**, 823-826.
- Emanuelsson,O., Nielsen,H., Brunak,S. and von Heijne,G. (2000) Predicting subcellular localization of proteins based

- on their N-terminal amino acid sequence. *J. Mol. Biol.* **300**, 1005-1016.
- Emanuelsson,O. (2002) Predicting protein subcellular localisation from amino acid sequence information. *Brief Bioinform.* **3**, 361-376.
- Fabbro,M. and Henderson,B.R. (2003) Regulation of tumor suppressors by nuclear-cytoplasmic shuttling. *Exp Cell Res.* **282**, 59-69.
- Gardy,J.L., Spencer,C., Wang,K., Ester,M., Tusnady,G.E., Simon,I., Hua,S., deFays,K., Lambert,C., Nakai,K. and Brinkman,F.S.L. (2003) PSORT-B, improving protein subcellular localization prediction for Gram-negative bacteria. *Nucleic Acids Res.* **31**, 3613-3617.
- Henderson,B.R. and Fagotto,F. (2002) The ins and outs of APC and beta-catenin nuclear transport. *EMBO Rep.* **3**, 834-839.
- Henikoff,S. and Henikoff,J.G. (1991) Automated assembly of protein blocks for database searching. *Nucleic Acids Research*, **19** 6565-6572.
- Hoetelmans,R., van Slooten,H.J., Keijzer,R., Erkeland,S. and van de Velde,C.J. (2000) Bcl-2 and Bax proteins are present in interphase nuclei of mammalian cells. *Cell Death Differ.* **7**, 384-392.
- Hsu,C.W. and Lin,C.J. (2002) A comparison on methods for multi-class support vector machines. *IEEE Trans. Neural Networks* **13**, 415-425.
- Jain,A.K., Duijn,R.P.W. and Mao,J. (2000) Statistical pattern recognition: a review. *IEEE Trans. Pattern Analysis and Machine Intelligence* **22**, 4-37.
- Joachims,T. (1999) Making large-scale SVM learning practical. In Schoelkopf B, Burges C, Smola,A. (eds.), *Advances in Kernel Methods - Support Vector Learning*, MIT-Press, Cambridge, USA.
- Kawashima,S., Ogata,H. and Kanehisa,M. (1999) AAindex: amino acid index database. *Nucleic Acids Res.* **27**, 368-369.
- Kittler,J.V., Hatef,M., Duijn R.P.W. and Matas,J. (1998) On combining classifiers. *IEEE Trans. Pattern Analysis and Machine Intelligence* **3**, 226-239.
- Kohonen,T. (1997) *Self-organizing Maps*.Springer-Verlag New York, Inc.,Secaucus, NJ
- Kohonen,T., Hynninen,J., Kangas,J. and Laaksonen,J. (1996) SOM_PAK: The Self-Organizing Map program package. Tech. Rep. A31, Helsinki University of Technology, Laboratory of Computer and Information Science, FIN-02150 Espoo, Finland.
- Kostiainen,T. and Lampinen,J., (2002) On the generative probability density model in the self-organizing map. *Neurocomputing* **48**, 217-228
- Lam,L. and Suen,C.Y. (1994) A theoretical analysis of the application of majority voting to pattern recognition. *Proceedings of Int. Conf. On Pattern Recognition* **1**, 418-420
- Lu,Z., Szafron,D., Greiner,R., Lu,P., Wishart,D.S., Poulin,B., Anvik,J., Macdonell,C. and Eisner,R. (2004) Predicting subcellular localization of proteins using machine-learned classifiers. *Bioinformatics* **20**, 547-556
- Luttrell,S.P. (1994) A Bayesian analysis of self-organizing maps. *Neural Computation* **6**, 767-794
- Mjolsness,E. and DeCoste,D. (2001) Machine Learning for Science: State of the Art and Future Prospects. *Science* **293**, 2051-2055
- Mohamed,E.M.M., Dick de,R., Robert,P.W.D. and Atalay,V. (2004) Almost autonomous training of mixtures of principal component analyzers. *Pat. Rec. Lett.* **25**, 1085-1095
- Mott,R., Schultz,J., Bork,P. and Ponting,C.P. (2002) Predicting protein cellular localization using a domain projection method. *Genome Res.* **12**, 1168-1174.
- Nair,R. and Rost,B. (2003) LOC3D: annotate sub-cellular localization for protein structures. *Nucleic Acids Res.* **31**, 3337-3340.
- Nakai,K. (2000) Protein sorting signals and prediction of subcellular localization. *Adv. Protein Chem.* **54**, 277-344.
- Noble,W.S. (2004) Support vector machine applications in computational biology. In Schoelkopf,B., Tsuda,K., Vert, J.P.(eds.), *Kernel Methods in Computational Biology*, pp 71-92. MIT Press, Cambridge, USA
- Özer,Ö.F., Özün,O., Tüzel,C.O., Atalay,V. and Cetin,A.E. (2001) Vision-based single-stroke character recognition for wearable computing. *IEEE Intel Sys.* **16**, 33-37
- Schmalz,D., Hucho,F. and Buchner,K. (1998) Nuclear import of protein kinase C occurs by a mechanism distinct from the mechanism used by proteins with a classical nuclear localization signal. *J Cell Sci.* **111**, 1823-1830
- Schoelkopf,C., Burges,J.C. and Smola,A.J. (1999) *Advances in Kernel Methods*. MIT Press.
- Scott,M.S., Thomas,D.Y. and Hallett,M.T. (2004) Predicting subcellular localization via protein motif co-occurrence. *Genome Res.* **14**, 1957-1966.
- Vapnik,V. (1998) *Statistical Learning Theory*. Wiley.
- Vesanto,J. and Alhoniemi,E. (2000) Clustering of the Self-Organizing Map. *IEEE Transactions on Neural Networks*, **11**, 586-600
- Wang,J., Delabie,J., Aasheim,H., Smeland,E., Myklebost,O., (2002) Clustering of the SOM easily reveals distinct gene expression patterns: results of a reanalysis of lymphoma study. *BMC Bioinformatics* **3**, 36